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Standardizing Selection Strengths to Study Selection in the Wild: A Critical Comparison and Suggestions for the Future

SHUICHI MATSUMURA, ROBERT ARLINGHAUS, AND ULF DIECKMANN

We critically review the main approaches for standardizing and comparing selection differentials and gradients among traits, populations, and species and assess their differential merits. In particular, we explain why the most widespread approach to standardizing selection, which measures selection differentials in units of a trait's phenotypic standard deviation, is less appropriate for characterizing the strengths of natural selection in wild populations. In contrast, an alternative standardization approach that uses a trait's phenotypic mean in addition to its standard deviation results in another dimensionless measure: the mean-standardized selection gradient. This measure offers some key benefits over alternative approaches, such as ease of interpretation, independence of a trait's variance, and the natural interplay of ecological and evolutionary dynamics, and yet it remains rarely used by evolutionary biologists. We explain how the more routine application of this measure will facilitate comparisons of selection strengths in the wild among traits, populations, and species.

Keywords: elasticity, natural selection, selection differential, selection gradient, standardization

Our world is filled with astonishingly diverse forms of organisms. Adaptation by natural selection to heterogeneous and changing environments is a critical driving force for bringing about this biodiversity (Darwin 1859). Phenotypes change according to both natural and anthropogenic selection pressures. Rapid phenotypic evolution resulting from human-induced environmental change, such as global warming or the harvesting of wild animals and plants, can be particularly troublesome, because such evolution may have undesirable consequences—for example, population declines, ecosystem instabilities, and forgone yield (Kinnison and Hendry 2001, Palumbi 2001, Jørgensen et al. 2007, Darimont et al. 2009). Anthropogenic selection pressures are likely to be very strong (Darimont et al. 2009); yet, their strength has been quantified and compared in few studies—for example, in the context of fisheries (Law 2007, Hutchings and Fraser 2008, Hard et al. 2008). To understand how various selective forces change the phenotypes of animals and plants in the wild, an important first step is to develop robust methods that quantify and compare the

power of natural and anthropogenic selection pressures acting on a diverse range of adaptive traits (Hendry 2005).

The general methodological framework for estimating the strength of selection was developed by Lande and colleagues in the late 1970s and early 1980s (e.g., Lande and Arnold 1983). Because of its conceptual appeal and ease of application, in a large number of empirical studies the strength of natural selection on adaptive traits has subsequently been estimated in a wide range of animals and plants using the regression-based methods pioneered by Lande and colleagues (for a review, see Kingsolver et al. 2001, Hereford et al. 2004). Their main innovation was to show how selection strength can be measured through a univariate or multivariate linear regression of relative fitness or components of fitness (such as survival at a particular life stage or the number of offspring; Arnold and Wade 1984) on the quantitative trait or traits of interest. Using this compelling approach, important insights have been generated into the mechanisms and patterns governing the direction and speed of evolutionary change, including ecoevolutionary feedback

Table 1. A sample of studies in which estimates of selection strengths were reported using the mean-standardized selection gradient.

Reference	Type of study	Subjects
Morgan and Schoen 1997	Empirical	Plants (floral characters in <i>Asclepias syriaca</i>)
Van Tienderen 2000	Illustrative	Hypothetical plants
Coulson et al. 2003	Empirical	Animals (neonatal traits in <i>Cervus elaphus</i>)
Hereford et al. 2004	Review of empirical studies	Estimates from 38 studies
Stinchcombe 2005	Empirical	Plants (a resistance trait and a size trait in <i>Ipomoea hederacea</i>)
Kelley et al. 2005	Empirical	Plants (defense traits in <i>Arabidopsis thaliana</i>)
Dibattista et al. 2007	Empirical	Animals (life-history traits including size and growth in <i>Negaprion brevirostris</i>)
Ehrlén and Münzbergová 2009	Empirical	Plants (floral characters in <i>Lathyrus vernus</i>)
Arlinghaus et al. 2009	Modeling	Animals (life-history traits in <i>Esox lucius</i>)
Mojica and Kelly 2010	Empirical	Plants (flower size in <i>Mimulus guttatus</i>)
Stinchcombe et al. 2010	Empirical	Plants (growth rate in <i>Impatiens capensis</i>)
Bolstad et al. 2010	Empirical	Plants (floral traits in <i>Dalechampia schottii</i>)
Simonsen and Stinchcombe 2010	Empirical	Plants (flowering time and size in <i>Ipomoea hederacea</i>)
Horvitz et al. 2010	Reanalysis of empirical data	Plants (<i>Calathea ovandensis</i>) and animals (<i>Cervus elaphus</i>)
Bromquist et al. 2011	Empirical	Animals (dominance rank in <i>Macaca mulatta</i>)
Reinhold 2011	Reanalysis of empirical data	Animals (acoustic signaling traits in insects and amphibians)

(Hendry 2005, Kingsolver and Pfennig 2007). The accumulation of data on selection pressures has, in turn, enabled researchers to compare selection strengths among different traits, populations, and species (e.g., Endler 1986, Kingsolver et al. 2001, Hereford et al. 2004, Hendry 2005, Kingsolver and Pfennig 2007). Yet, the utility of such comparisons is still crucially limited by two fundamental problems. First, because different phenotypic traits have different units, any direct comparison of selection strengths among them is limited. Second, reported indicators of selection strength are often confounded by population- or trait-specific properties, such as a population's generation time or a trait's heritability, phenotypic variance, or phenotypic mean. These two problems hamper a complete understanding of selection responses of different traits in different populations experiencing different selective environments. To enable meaningful comparisons, selection strengths have to be adequately standardized in a way that ensures that all confounding factors are removed.

The objective of this article is to review the differential merits of alternative ways of standardizing selection strengths. In particular, we highlight the important advantages of the mean-standardized measure, which render this approach the most useful for comparing selection strengths in wild populations of animals and plants. Complementing the commendable pioneering discussion by Hereford and colleagues (2004; table 1), our exposition is meant to provide

an easily accessible one-stop overview of these matters for the benefit of all those interested in comparing the strengths of selection in natural populations. We also provide a worked-out example and an overview of the main steps to show how to obtain and standardize measures of selection strength from empirical data.

Selection differentials, responses, and gradients

One of the central measures of selection in traditional quantitative genetics and artificial breeding is the selection differential, S , which measures the mean phenotypic value of individuals selected as parents, expressed relative to the mean phenotypic value of all individuals in the parental generation before selection (Falconer and Mackay 1996). In selection experiments or artificial breeding efforts, selection often operates through truncation, by admitting for reproduction only individuals whose phenotypic trait values exceed a given threshold (figure 1a). A form of viability selection, such truncation selection is recognized as the most

efficient implementation of artificial directional selection (Crow and Kimura 1979). By contrast, truncation selection rarely occurs in natural populations, because there is usually no threshold phenotypic value that would distinguish individuals that reproduce—or survive—from those that do not. For natural selection, instead, the selection differential is obtained as the phenotypic mean of individuals weighted by their fitness, expressed relative to the population's phenotypic mean, in which all individuals are weighted alike (figure 1c; see also the illustrative example in box 1). The empirical measurement of selection differentials therefore requires an estimation of the differential fitness of individuals, depending on the phenotypic traits characterizing those individuals. This is often difficult—or even practically impossible—especially when a trait's fitness consequences are distributed over an organism's full life cycle. Therefore, selection differentials are frequently calculated on the basis of a fitness component (Arnold and Wade 1984)—for example, survival at a particular life stage or the number of offspring produced over a certain period (e.g., a year). Such differentials more narrowly measure the strength of selection acting on an adaptive trait at a particular stage of the life cycle.

A more readily observable measure of selection strength is the selection response, R , which quantifies the amount of evolutionary (i.e., genetic) change in a phenotypic trait value between generations and is therefore defined as the

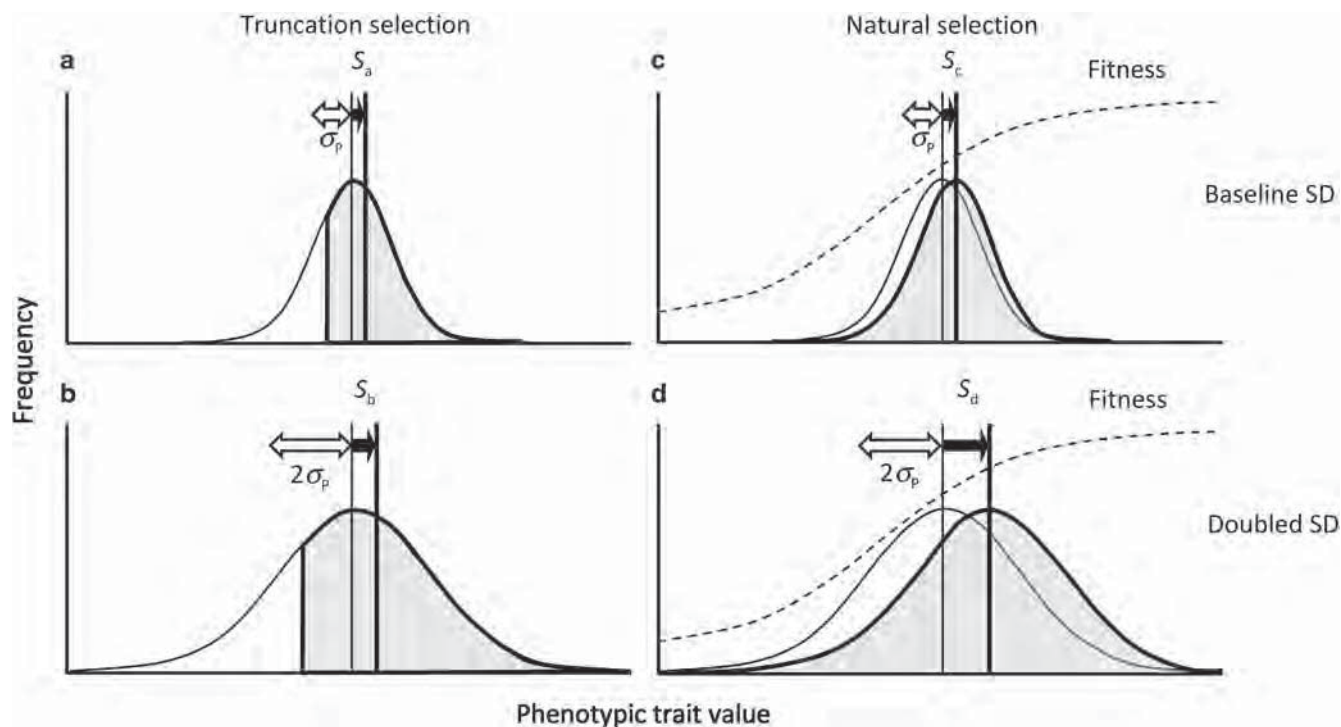


Figure 1. Schematic illustration of how a selection differential S is defined as the shift in a trait's phenotypic mean before and after selection. The two columns show this for truncation selection (left) and natural selection (right). The two rows show how S is altered by different degrees of phenotypic variability, here illustrated by two normal distributions, one of which has twice the standard deviation (doubled SD, $2\sigma_p$; bottom) of the other (baseline SD, σ_p ; top). Under truncation selection (panels (a) and (b)), S is the difference between the phenotypic mean μ_p of all individuals in a population (the thin line) and the phenotypic mean of those individuals that are selected as parents for reproduction (the thick line). When the same proportion of individuals is selected in the baseline population (a) and in the population with double SD (b), the selection differential is twice as large as S_a ($S_b = 2S_a$). This makes it natural to consider the SD-standardized selection gradient ($\beta_\sigma = S/\sigma_p$), which conveniently measures the proportion of individuals of the whole population that are selected as parents for reproduction. This gives $\beta_{\sigma,b} = S_b/2\sigma_p = 2S_a/2\sigma_p = S_a/\sigma_p = \beta_{\sigma,a}$, which reflects that truncation selection was applied in the same way (i.e., is described by the same truncation proportion) in both populations. Under natural selection (panels (c) and (d)), S is the difference between the mean of a population's phenotypic distribution (the thin continuous line) and of this distribution weighted with the differential fitness of phenotypes (the thin dashed line), resulting in the product shown by the thick line. When the same fitness function acts on the baseline population (c) and the population with double SD (d), the selection differential is (approximately) four times as large as S_c ($S_d = 4S_c$). This makes it natural to consider the mean-standardized selection gradient $\beta_\mu = \mu_p S/\sigma_p^2$, which conveniently measures the elasticity of fitness. This gives $\beta_{\mu,d} = \mu_p S_d/(2\sigma_p)^2 = \mu_p(4S_c)/(4\sigma_p^2) = \mu_p S_c/\sigma_p^2 = \beta_{\mu,c}$, which reflects that natural selection applied in the same way (i.e., is described by the same fitness function) in both populations.

change in a population's mean phenotypic value during one generation. In traditional quantitative genetics, the selection response is associated with the selection differential according to the breeder's equation,

$$R = h^2 S, \quad (1)$$

where $h^2 = \sigma_A^2/\sigma_p^2$ is the narrow-sense heritability, and σ_A^2 and σ_p^2 are a population's additive genetic variance and its phenotypic variance, respectively (Falconer and Mackay 1996). It is therefore evident that selection responses depend

on population- and trait-specific properties (h^2 , or σ_A^2 and σ_p^2). As we will see further below in this section, an analogous conclusion applies to selection differentials, even though this is less commonly emphasized.

The methods developed by Lande and his coauthors (e.g., Lande and Arnold 1983) measure the selection gradient, β , which is the average slope (or gradient) of a trait's fitness landscape experienced by a population in a given ecological environment. Such a fitness landscape represents how the fitness of individuals changes with a change in their trait value. It has been shown that β can be estimated as the

Box 1. An illustrative example for estimating mean-standardized selection gradients using model-based analyses of wild populations.

To illustrate the usefulness of the mean-standardized selection gradient, here, we show how to model selection differentials in a hypothetical semelparous anadromous fish species, such as a salmonid. We hope this example will help theoretical ecologists wishing to apply the mean-standardized selection gradient to their own models.

Model description

We consider a simple fish life history with individuals that are born in a river, migrate to the ocean at an age of 1 year, grow in the ocean, return to the river at an age of a_m years, reproduce, and die after reproduction. We examine the date of river entry, d , and the annual juvenile growth increment, h , as evolving traits.

The success of spawning migration in salmonids has been reported to depend on river temperature, with the optimal temperature differing between populations (e.g., Farrell et al. 2008). Any increase in river temperature—for example, due to global warming—is therefore expected to cause selection pressures on the date of river entry. Here, we simply assume that river temperature decreases linearly from late summer to autumn. For fish entering the river at day d , the deviation from the optimal temperature is given by $T_{\text{dev}}(d) = -b_1(d - d_{\text{opt}}) + T$, where d_{opt} is the optimal date of river entry without climate change, $b_1 > 0$ is the rate of seasonal temperature change, and T measures the increase of river temperature due to climate change. The survival probability during the spawning migration from the river mouth to the spawning site is $s_{\text{mig}} = b_2 \exp(-b_3(T_{\text{dev}}(d)/T_u)^2)$, where b_2 is the survival probability at the optimal temperature, $b_3 > 0$ determines the temperature sensitivity of migration survival, and T_u is a unit-standardizing constant.

The growth trajectory of individuals is described using the biphasic somatic growth model of Lester and colleagues (2004): The length at age a_m is represented as

$$L_{a_m} = \frac{3}{g+3}(L_{a_m-1} + h) = \frac{3a_m h}{g+3},$$

where h is the annual juvenile growth increment and g is the annual reproductive investment. The fecundity f_{a_m} of fish at age a_m that have successfully arrived at spawning sites is allometrically given by $f_{a_m} = b_4 g (L_{a_m}/L_u)^{b_5}$, where b_4 and b_5 are the allometric parameters and L_u is a unit-standardizing constant. The basic annual natural mortality is given by $m = h/h_{\text{max}}$, where h_{max} is the maximum growth capacity beyond which survival has a probability of zero (Matsumura et al. 2011). The annual survival probability of hatched larvae (newborn survival, s_0) and fish in the ocean (juvenile survival, s_j) is then represented as $s_0 = b_6(1 - m)$ and $s_j = b_7(1 - m)$, respectively, where b_6 and b_7 are parameters.

We assume a polymorphic resident population with trait value x being normally distributed around the population mean μ_p with phenotypic variance σ_p^2 . The dynamics of fish with trait values x in this population is described by a population projection matrix:

$$\mathbf{A}(x) = \begin{pmatrix} 0 & 0 & \cdots & 0 & s_0(x)f_{a_m}(x)s_{\text{mig}}(x) \\ s_j(x) & 0 & \cdots & 0 & 0 \\ 0 & s_j(x) & \cdots & 0 & 0 \\ \vdots & \vdots & \ddots & \vdots & \vdots \\ 0 & 0 & \cdots & s_j(x) & 0 \end{pmatrix}.$$

Changes in the age structure and density of fish with trait values x are therefore described by $\mathbf{N}'(x) = \mathbf{A}(x)\mathbf{N}(x)$, where the vectors $\mathbf{N}(x) = (N_1(x), N_2(x), \dots, N_{a_m}(x))^T$ and $\mathbf{N}'(x) = (N'_1(x), N'_2(x), \dots, N'_{a_m}(x))^T$ contain the densities of fish across all age classes in years t and $t + 1$, respectively.

Calculating selection differentials

The dominant eigenvalue, $\lambda(x)$, of the population projection matrix, $\mathbf{A}(x)$, of trait value x is known to represent the annual population growth rate, which is commonly used to measure trait-specific lifetime fitness (e.g., Metz et al. 1992, Arlinghaus et al. 2009). The selection differential per year is then calculated as $S = \int x\lambda(x)p(x)dx / \int \lambda(x)p(x)dx - \mu_p$ (figure 1), where $\lambda(x)$ and $p(x)$ are the fitness and probability density of trait value x , respectively. The first term in this difference corresponds to the average phenotypic value of the population after selection—that is, the average phenotypic value of parents of the next generation (Arlinghaus et al. 2009).

In the present example, we consider the effects of an increase T in river temperature (e.g., due to global warming), so that the fitness of fish that enter the river later than at their traditional optimal date is expected to be raised. In other words, the proportion of latecomers will be larger in the spawning population than in the original population, which makes the first term in the above difference larger than the second, which, in turn, results in a positive selection differential.

Box 1. continued.

The influence of trait variance on the three standardized measures of selection strength

Using this example, we first examine how three standardized measures of selection strength change for different levels of the population's phenotypic variability. We focus on the date of river entry (d) and calculate the selection differential (S) for d caused by the considered increase in river temperature. We vary the phenotypic standard deviation (σ_p^2) and examine the resultant changes of the three standardized measures under fixed strengths of selection (figure 2). It is easily seen that only the mean-standardized selection gradient is approximately independent of phenotypic variability.

Merging evolution and ecology through elasticity path diagrams

Next, we illustrate the potential of the elasticity path diagram (figure 3; van Tienderen 2000) to merge evolution and ecology and to facilitate comparisons across traits and subpopulations. We consider two hypothetical endangered fish subpopulations that live in different habitats and differ in their life-history patterns (subpopulations A and B). Considerable heritable variation in the age of maturation has been reported in salmonids (e.g., Friars and Smith 2010), so we assume that the ages of fish returning to the river and reproducing in subpopulation A and B are $a_m = 4$ years and $a_m = 2$ years, respectively. We also assume that the basic annual natural mortality increases less rapidly with the growth increment in subpopulation A ($h_{\max} = 60$ centimeters [cm]) than in subpopulation B ($h_{\max} = 55$ cm).

In subpopulation A, the fitness elasticity of juvenile survival (0.75) is larger than that of any of the other vital rates (0.25). This has an implication for population managers: Improving juvenile survival would be a good strategy to increase the population growth rate of subpopulation A. At the same time, it is predicted that natural selection would favor traits improving juvenile survival. Although smaller growth increments lead to higher juvenile survival in our model, this also reduces fecundity. Consequently, selection on growth in subpopulation A is weaker than that on the date of river entry. By contrast, in subpopulation B, the fitness elasticity of juvenile survival is not larger than those of the other vital rates, and the selection acting on growth is stronger than that on the date of river entry. Therefore, by providing ecological and evolutionary measures as mean-standardized values and considering them together, the elasticity path diagram facilitates comparisons across traits, subpopulations, and species in natural populations. The extrapolation of estimates from a well-known subpopulation or species to others is also enabled by using such mean-standardized values, although this must always be done with care, because all empirically estimated elasticities are dependent on the considered ecological environments.

coefficient of a linear regression of the relative fitness of a phenotypic trait—or, more generally, as a partial regression coefficient in the case of multivariate traits (Lande 1979, Lande and Arnold 1983). In this framework, the selection differential is the covariance between the phenotypic trait and fitness, whereas the selection gradient is the covariance divided by the variance of the phenotypic trait. Generally, β , S , and R are related as in equation 2,

$$\beta = S / \sigma_p^2 = R / \sigma_A^2. \quad (2)$$

Because β measures the sensitivity of fitness to changes in a trait's value, it is a more direct measure of selection strength in natural populations than is S or R . From equation 2, we see that the selection differential,

$$S = \sigma_p^2 \beta, \quad (3)$$

confounds the selection gradient with a population's phenotypic variance, which means that, just like R , S is not an ideal measure for comparing selection strengths among different traits, populations, or species.

In multivariate cases, selection is acting on n different traits, and equation 2 generalizes to

$$R = G\beta, \quad (4)$$

where $R = (R_1, R_2, \dots, R_n)^T$ and $\beta = (\beta_1, \beta_2, \dots, \beta_n)^T$ denote the vectors of multivariate selection responses and selection gradients, respectively. The $n \times n$ matrix G is called the *additive genetic variance-covariance matrix* and comprises the additive genetic variances of all n traits, as well as the covariances for all $n(n-1)$ pairs of different traits (Lande and Arnold 1983). In this manner, equation 4 captures both the direct evolutionary trait changes resulting from selection on a given focal trait and the indirect trait changes resulting from selection on other traits that are genetically correlated with the focal trait (for a more detailed discussion of multivariate cases, see Hansen and Houle [2008]). In the present article, we will focus on univariate selection, because our main interest is not in selection responses but in selection differentials and gradients for particular traits.

Standardizing selection strength

When researchers equipped with the aforementioned concepts, methods, and equations strive to compare selection strengths among different traits, a fundamental problem arises: Neither the selection differential, nor the selection response, nor the selection gradient is dimensionless. If the trait of interest is measured in kilograms (kg), for instance, the selection differential, selection response, and selection gradient will have units of kg, kg, and kg^{-1} , respectively. We illustrate the resultant difficulties by focusing on the

selection differential, the most traditional measure of these three. Let us consider an example from the literature on fisheries-induced evolution, which has attracted considerable attention in recent years (e.g., Jørgensen et al. 2007, Kuparinen and Merilä 2007): Hard and colleagues (2008) reviewed empirical estimates of selection differentials imposed by fishing and reported selection differentials on body length in Atlantic cod (*Gadus morhua*; S ranged from -1 to 2 centimeters [cm]), on body weight in Atlantic salmon (*Salmo salar*; S ranged from -0.08 to -0.52 kg), and on body girth in sockeye salmon (*Oncorhynchus nerka*; S ranged from -0.6 to -3.6 millimeters). From an absolute comparison of these values, there is no way that one could tell which trait is under the strongest selection. Therefore, it is evident that S —and, analogously, R and β —must be standardized before any attempted comparison of selection strengths.

There are two obvious approaches to this standardization. First, selection differentials can be measured in units of the phenotypic mean of the trait under selection by considering the ratio S/μ_p . Representing in this manner a trait's proportional change relative to its mean is probably the most intuitive approach to standardization. Indeed, when the selection response is measured in units of the phenotypic mean, R/μ_p , the resultant ratio usefully describes the proportional change in the trait value from one generation to the next (Roff 2002). Interestingly, this type of standardization is rarely reported for selection differentials. Alternatively, selection differentials can be expressed in units of the phenotypic standard deviation (SD) of the trait under selection by considering the ratio S/σ_p . As we will explain below, this currently most popular approach to standardizing selection strengths originated in the context of controlled selection experiments typical in artificial breeding studies of animals and plants but, unfortunately, is not optimally suited to standardizing the strength of natural selection.

Recognition of a third standardized measure is a relatively recent development, because this additional measure is not as obviously constructed as the two described so far. Specifically, the selection differential can be standardized by jointly using the phenotypic mean and the phenotypic variance of the trait under selection, considering the ratio $\mu_p S/\sigma_p^2$. The motivation for this particular—at first sight, seemingly odd—construction is to remove the phenotypic variance's confounding effect on the selection differential (equation 3) and yet obtain a dimensionless measure.

As was already explained above, the selection gradient measures the sensitivity of fitness to changes in a trait's value and is therefore considered a more direct measure of selection strength than is the selection differential. According to equation 2, the second and third standardized measures of selection strength are related to the selection gradient as equations 5 and 6, respectively. We can therefore see that these measures can be interpreted as a fitness landscape's slope measured in units of, respectively, the phenotypic SD, σ_p , or the phenotypic mean, μ_p . For this reason, the two

measures were termed the *variance-standardized selection gradient* (β_σ) and the *mean-standardized selection gradient* (β_μ) by Hereford and colleagues (2004):

$$\beta_\sigma = S/\sigma_p = \sigma_p\beta, \quad (5)$$

$$\beta_\mu = \mu_p S/\sigma_p^2 = \mu_p\beta. \quad (6)$$

In the present article, we call the former measure the *SD-standardized selection gradient*, because it actually is standardized by the phenotypic SD and not by the phenotypic variance.

Standardizing selection strength by the phenotypic SD

The SD-standardized selection gradient (equation 5) is also known as the *selection intensity* (or the *intensity of selection*) in traditional quantitative genetics (table 2; Falconer 1960, Bulmer 1980, Falconer and Mackay 1996, Roff 1997). In the context of truncation selection, the selection intensity has a particularly compelling interpretation, because it then directly measures the proportion of individuals that are not selected for reproduction. If this proportion increases, the selection intensity increases. What is important for comparative purposes, especially in artificial breeding efforts, is that the selection intensity is independent of the variability of the trait considered. If the selected proportion is the same, the selection intensity will be the same, regardless of the variability of the trait (figure 1a, 1b; Falconer and Mackay 1996, Roff 1997). Because of its utility for describing truncation selection, the selection intensity has long been considered a good measure of selection strength in general (Falconer and Mackay 1996).

This tradition of standardizing selection differentials by phenotypic SDs has continued even after measurements of selection strength were extended to natural populations. When Lande and Arnold (1983) applied their newly developed regression method to empirical data, they acknowledged the established tradition by explaining that the “observed selection differential on a particular character is often expressed in terms of phenotypic standard deviation” (p. 1219). Since then, the SD-standardized selection gradient has been widely used in comparative studies on natural selection in the wild. For example, Kingsolver and colleagues (Kingsolver et al. 2001, Hoekstra et al. 2001, Kingsolver and Diamond 2011) reviewed an extensive body of empirical studies and conducted a meta-analysis of selection strengths in natural populations of animals and plants using the SD-standardized selection gradient.

Even outside the context of truncation selection, the SD-standardized selection gradient has a simple interpretation: It measures the change in relative fitness that results from increasing the trait value by 1 phenotypic SD. For example, for $\beta_\sigma = 0.4$, shifting trait values up by 1 SD from their phenotypic mean increases relative fitness by 40%. (One must keep in mind, however, that this interpretation

Table 2. Alternative terms used for the two major standardized measures of selection strength across a sample of scientific publications.

Measure	Term	Reference
$\beta_\sigma = S/\sigma_p = \sigma_p\beta$	Selection intensity (intensity of selection)	Falconer 1960 and many others
	Standardized selection gradient	Lande and Arnold 1983 and many others
	Variance-standardized selection gradient	Hereford et al. 2004
	Standard-deviation (SD)-standardized selection differential	Arlinghaus et al. 2009
	Selection gradient in units of the phenotypic standard deviation	The present study (recommended long form)
$\beta_\mu = \mu_p S/\sigma_p^2 = \mu_p\beta$	SD-standardized selection gradient	The present study (recommended short form)
	Proportional sensitivity ^a	Caswell et al. 1978
	Elasticity ^a	De Kroon et al. 1986
	Elasticity of selection	Morgan and Schoen 1997
	Mean-standardized selection gradient	Hansen et al. 2003, Hereford et al. 2004
	Fitness elasticity	Stinchcombe 2005
	Mean-standardized selection coefficient	Dibattista et al. 2007
	Mean-and-variance-standardized selection differential	Arlinghaus et al. 2009
	Selection gradient in units of the phenotypic mean	The present study (recommended long form)
	Mean-standardized selection gradient	The present study (recommended short form)

^aDefined for elements of a population's projection matrix in the context of matrix models of population dynamics.

is only permissible when selection is frequency independent or when the fraction of individuals whose trait values are shifted is negligible relative to the considered population as a whole, because, otherwise, the trait shift may reshape the fitness landscape.) Unfortunately, this intuitive interpretation does not reflect the fact that the SD-standardized selection gradient remains confounded by the variance of the focal trait under selection and, therefore, by an idiosyncratic property of the selected population. We will explain and discuss this limitation further below.

Standardizing selection strength by the phenotypic mean

Attention to the mean-standardized selection gradient (equation 6) has recently increased. Mean-standardized selection responses were already reported by Johnson and colleagues (1955). To our knowledge, Morgan and Schoen (1997) may have been the first to give systematic consideration to the specific advantages of using the mean-standardized selection gradient in quantitative genetics. Further discussion about the prospects of mean-standardized selection gradients can be found in van Tienderen (2000) and Hereford and colleagues (2004). In the latter study, the database of Kingsolver

and colleagues (2001) was reanalyzed using the mean-standardized measure, and it was concluded that it was superior to the SD-standardized measure when selection strength on different traits was to be compared in natural populations. To understand this claim, it is helpful to look at mean-standardized measures in the context of the breeder's equation (equation 1). Using equation 2 and applying mean standardization not only to the selection differential but also to all other variables, equation 1 can be rewritten as

$$R/\mu_p = (\sigma_A/\mu_p)^2 (\mu_p\beta) \text{ or } r = a^2\beta_\mu, \quad (7)$$

where the proportional selection response, r (where $r = R/\mu_p$), is predicted by the mean-standardized selection gradient, β_μ , and the coefficient of genetic variation, a (where $a = \sigma_A/\mu_p$). The latter has been suggested as a suitable measure of evolvability (Houle 1992, Hansen et al. 2003), because it characterizes a population's ability to respond to selection. This can easily be understood because a larger genetic variation (SD, σ_A) relative to a small phenotypic mean (μ_p) implies a higher potential for evolution and, therefore, evolvability. In equation 7, all variables

are dimensionless and are measured in units of the trait's phenotypic mean. As a result, selection strength (β_μ) and evolvability (a) are cleanly separated. On this basis, selection strengths, evolvabilities, and proportional selection responses can consistently be compared among different traits, populations, and species (Hansen et al. 2003, Hereford et al. 2004).

Analogous to the SD-standardized selection gradient (β_σ), the mean-standardized selection gradient (β_μ) has a simple interpretation: It measures the change in relative fitness that results from doubling the trait value. For example, for $\beta_\mu = 0.4$, shifting trait values up by 100% increases relative fitness by 40% (the aforementioned caveat about potential changes in the fitness landscape resulting from such a shift of course applies here, too).

Which measure of selection strength is more suitable for natural populations?

In their meta-analyses of selection strengths in natural populations, Kingsolver and colleagues (2001) and Hereford and colleagues (2004) investigated similar data sets but used different standardizations of selection strength and thereby came to qualitatively different conclusions. In the former

Box 2. Natural scales and their implications.

In many contexts, a trait's natural scale would be given by its phenotypic standard deviation (SD). In the context of natural selection, however, this SD is disqualified from serving in this role, since a trait's phenotypic variance is of such central dynamical importance for scaling a trait's selection differential and response (figure 2). Because the phenotypic variance is needed for removing this confounding effect of phenotypic variability from selection differentials, it is unsuitable for rendering selection gradients dimensionless. In most circumstances, the most natural scale that remains is given by a trait's phenotypic mean. This is ultimately because the majority of traits have a natural minimum—typically designated by a trait value of 0. The difference between this minimum and a trait's current phenotypic mean establishes a natural scale suitable for standardization.

Against this background, we can now systematically appreciate two exceptions mentioned in the text. First, when a quantitative trait possesses both a natural minimum and a natural maximum, the two natural scales given by the difference between the trait's current phenotypic mean and its minimum or maximum, respectively, can alternatively be employed for standardization. For such traits, however, it will often be more natural simply to use the difference between the trait's minimum and maximum as the natural scale for standardization, which overcomes the ambiguity that would arise otherwise. For example, Stinchcombe (2005) considered two phenotypic traits of a plant species—resistance and susceptibility to herbivore damage, which both vary between 0 and 1 and are direct complements to each other by always adding up to 1—and pointed out the problem of obtaining different values for the mean-standardized selection gradient of these traits. Here, we first argue that this problem can be avoided altogether by using the difference between the traits' minimum (0) and its maximum (1) as the natural scale for standardization (in this example, the difference is 1). Second, when a quantitative trait has no natural minimum or maximum or when the values of these quantities are highly uncertain, the implied natural scales do not exist or are empirically unreliable and, as such, should not be used for standardization.

More fundamentally, whether a variable can be standardized is related to measurement theory: Houle and colleagues (2011) claimed that permissible transformations differ depending on a variable's scale type, such as nominal or ordinal numbers, intervals, or ratios. Another important issue pointed out by Houle and colleagues (2011) is the importance of concepts and hypotheses for measurement: Different contexts assign different meanings (and sometimes different scale types) to the same measurement.

study, it was concluded that natural selection was generally not strong, whereas in the latter study, the opposite was suggested. With such fundamental conclusions at stake, the choice of standardization is certainly no trifling matter. The question therefore naturally arises of which of the two standardized measures— β_{σ} or β_{μ} —can provide the more relevant assessment of selection strength in comparative studies of natural selection. To address this question, below, we appraise three limitations and three advantages of using the mean-standardized selection gradient, β_{μ} .

Three limitations of the mean-standardized selection gradient were pointed out in recent reviews and meta-analyses on selection strength (Kingsolver and Pfennig 2007, Kingsolver and Diamond 2011), which led the authors of those studies to express their preference for using the SD-standardized selection gradient. First, mean-standardized selection gradients should not be applied to traits whose scale lacks a natural origin. If the origin of a trait's scale is arbitrary, different researchers will make different choices, and the effect of these arbitrary choices on the phenotypic mean will therefore contaminate the resultant mean-standardized selection gradients. A related point was raised by Stinchcombe (2005), who highlighted that mean-standardized selection gradients are problematic when they are applied to fractions that vary between 0 and 1. Although such a trait, f , and its complement, $1 - f$, necessarily experience selection of equal strength (but in opposite directions), the two corresponding mean-standardized selection gradients differ not only in sign but also in magnitude (because f and $1 - f$ generally have

different means), so it is not self-evident which of these two magnitudes to report for the mean-standardized selection gradient. We further discuss and try to resolve this point in box 2. Second, the information needed to compute mean-standardized selection gradients is not always reported in publications. After Lande and Arnold's (1983) influential work, SD-standardized values were reported in many studies, but phenotypic means were often not documented in the corresponding publications. Because of this historical bias, the scope for meta-analyses of earlier studies of selection strengths in natural populations is narrower for the mean-standardized than for the SD-standardized selection gradient. Although the last statement seems incontrovertible, we suggest that this historical and practical limitation must not outweigh the more conceptual arguments that we discuss elsewhere in the present article. Third, larger mean-standardized selection gradients appear to occur in populations with smaller phenotypic coefficients of variation. Further investigations will be needed to test this correlation and, if it is corroborated as robust, to determine the underlying reasons.

We now turn to describing three key advantages of using mean-standardized selection gradients for comparing selection strengths in natural populations. First, these measures are easy to interpret in terms of fitness elasticities. Second, they are not confounded by a population's phenotypic variability. Third, they facilitate the liaison of ecological and evolutionary studies. Although both Kingsolver and Pfennig (2007) and Stinchcombe (2005) acknowledged the first

advantage, neither mentioned the second or third. After describing the first of these advantages briefly, we discuss the second in greater detail before explaining the third toward the end of this section.

First advantage: Interpretation as elasticities. As we described above, the mean-standardized selection gradient measures the change in relative fitness that results from doubling the trait value. More generally, increasing the trait value by a fraction (f) of the mean trait value causes relative fitness to increase by $\beta_\mu f$. Therefore, β_μ measures fitness elasticities, which is why this measure has also been called the *elasticity of selection* (Morgan and Schoen 1997).

Hereford and colleagues (2004) pointed out that one of the advantages of using mean-standardized selection gradients, stemming from their interpretation as elasticities, is the existence of a useful benchmark for relatively strong selection: When the selected trait is fitness, $\beta_\mu = 1$. Therefore, a mean-standardized selection gradient of $|\beta_\mu| > 1$ describes a fitness landscape with a relatively steep slope around the population's phenotypic mean (Hereford et al. 2004), because a change in trait value then results in a substantial change in fitness, which in turn implies relatively strong selection. We have noted that some researchers were suspicious about the fact that the elasticity of a trait or fitness component can exceed the elasticity of fitness itself. To escape a pitfall in understanding, it is worth recalling that there is no biological reason that would generally limit the steepness of a fitness landscape's slope and, in particular, would prevent this slope from exceeding the inverse ($1/\mu_p$) of a population's phenotypic mean. For example, $\beta_\mu = 1.2$ means that shifting trait values up by 100% increases relative fitness by 120%, which just shows that fitness is very sensitive to a change in that particular trait. Such high sensitivities are expected, for instance, in traits that rather abruptly enable survival or reproduction when exceeding a certain threshold.

Second advantage: Independence of a population's phenotypic variability. To examine the second advantage, we carried out an illustrative model-based analysis in which the three standardized measures of selection strength were tested (for a description of the underlying model, see box 1). We varied the variability of a phenotypic trait that was subject to selection of fixed strength (figure 1d). The results demonstrate that the SD-standardized selection gradient and the proportional selection differential are not invariant; instead, they appear to increase linearly and quadratically with σ_p , respectively (figure 2a, 2b). In contrast, the mean-standardized selection gradient remains almost constant under such variation, as long as the phenotypic SD is not too large (figure 2c).

These features are readily understood by examining the relationship between the selection gradient and the fitness landscape, $\lambda(x)$, of a phenotypic trait x . Assuming a normal distribution of trait values around the phenotypic mean (μ_p)

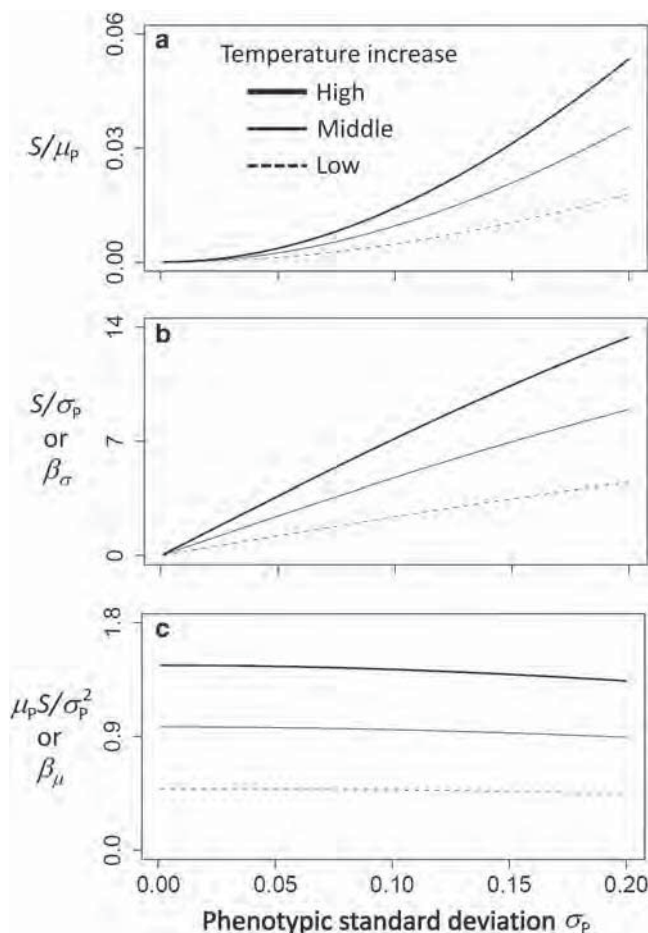


Figure 2. Dependence of three alternative standardized measures of selection strength on a population's phenotypic variability. The panels show absolute values of (a) S/μ_p , (b) $\beta_\sigma = S/\sigma_p = \sigma_p \beta$, and (c) $\beta_\mu = \mu_p S/\sigma_p^2 = \mu_p \beta$, respectively. Three different levels of river temperature are considered (low, middle, and high), which result in selection of increasing strength (see box 1 for model details). Parameters: $a_m = 2$ years, $d_{opt} = 50$ d, $g = 0.5$, $h = 25$ centimeters (cm) per year, $h_{max} = 55$ cm per year, $T = 1$ degrees Celsius ($^{\circ}\text{C}$), $T_u = 1^{\circ}\text{C}$, $L_u = 1$ cm, $b_1 = 0.1^{\circ}\text{C}$ per day, $b_2 = 0.9$, $b_3 = 0.1$, $b_4 = 0.05$, $b_5 = 3$, $b_6 = 0.00032$, and $b_7 = 0.8$.

with variance σ_p^2 , and expanding $\lambda(x)$ around μ_p , Phillips and Arnold (1989) showed that

$$\beta = S/\sigma_p^2 = \left(\frac{d\lambda}{dx} + \frac{1}{2} \frac{d^3\lambda}{dx^3} \sigma_p^2 + \frac{1}{8} \frac{d^5\lambda}{dx^5} \sigma_p^4 + \dots \right)_{x=\mu_p} \approx \left(\frac{d\lambda}{dx} \right)_{x=\mu_p}. \quad (8)$$

Since the higher-order terms are expected to be negligible when σ_p is sufficiently small, the selection gradient (β) approximately equals the slope ($d\lambda/dx$) of the fitness landscape at the phenotypic mean. From this, we immediately see that the mean-standardized selection gradient ($\beta_\mu = \mu_p \beta \approx \mu_p (d\lambda/dx)_{x=\mu_p}$) is not only

dimensionless but also almost constant under variation of σ_p (as long as σ_p is small) and approximates the fitness landscape's slope around the trait's mean in units of the latter. By contrast, the SD-standardized selection gradient ($\beta_\sigma = \sigma_p \beta \approx \sigma_p (d\lambda/dx)_{x=\mu_p}$) and the selection intensity, or the proportional selection differential ($S/\mu_p = \sigma_p^2 \beta / \mu_p \approx \sigma_p^2 (d\lambda/dx)_{x=\mu_p} / \mu_p$), are shown to increase approximately linearly and quadratically with σ_p , respectively.

These results highlight a major disadvantage of the SD-standardized selection gradient (β_σ) as a measure of selection strength. Even when selection pressures and phenotypic means are identical, populations with different degrees of phenotypic variability will exhibit different values of β_σ on the same trait of interest. In other words, the SD-standardized selection gradient varies with a population's ecological state, since a population's phenotypic variance is readily affected, for example, by whether it experiences a wide or narrow range of environmental conditions. Consequently, β_σ is not a pure measure of selection strength but, instead, confounds this with a population's phenotypic variability in the considered trait. The benefit of standardization by the phenotypic SD in the context of truncation selection—that is, its independence of the phenotypic variability of the trait—is therefore lost in the context of natural selection. This is because artificial truncation selection often restricts breeding to a specific fraction of individuals, whereas this is almost never the case under natural conditions. By contrast, the mean-standardized selection gradient is largely independent of a trait's phenotypic variability. Therefore, mean-standardized selection gradients are fundamentally superior for comparing the strengths of natural selection among traits, populations, and species across different ecological environments.

Some may dismiss the utility of the mean-standardized selection gradient because it varies with the phenotypic mean of a trait instead of with its SD even when the slope of a fitness landscape is kept constant. In our assessment, however, this feature further underscores the utility of the mean-standardized selection gradient, because it reflects a real biological difference. To see this, imagine a population of guppies (*Poecilia reticulata*; with a mean length of 5 cm) and a population of white sturgeon (*Acipenser transmontanus*; with a mean length of 500 cm). When selection on length is described in these two populations by fitness landscapes that have the same slopes of $\beta = 0.01 \text{ cm}^{-1}$ around the corresponding phenotypic means, the relative fitness of a 6-cm-long guppy (which is 20% larger than its population's phenotypic mean) is $1.01 = 1 + (6 - 5) \times 0.01$, whereas that of a 600-cm-long sturgeon (which is 20% larger than its population's phenotypic mean) is $2 = 1 + (600 - 500) \times 0.01$. It would presumably not occur to a field biologist to claim that length selection in these two populations was of equal strength. This intuitive understanding is well reflected when comparing β_μ values (which are $5 \times 0.01 = 0.05$ for guppies and $500 \times 0.01 = 5$ for white sturgeon) but not when comparing β values directly (which are identical for

guppies and sturgeon). In other words, the same slope of the fitness landscape will commonly be interpreted as representing different selection strengths depending on the considered trait's phenotypic mean. Therefore, the mean-standardized selection gradient—that is, the slope of a fitness landscape at a population's phenotypic mean measured in units of the latter—often serves as a biologically more appropriate measure of selection strength than the slope itself. In conjunction with the coefficient of genetic variation divided by the phenotypic mean—that is, evolvability (σ_A/μ_p)—it is also a good predictor of the proportional selection response (equation 3).

Third advantage: Merging ecology and evolution. We conclude this section by highlighting a third fundamental advantage associated with the use of the mean-standardized selection gradient: It naturally merges ecological and evolutionary dynamics through its decomposition into constituent components. Figure 3a is a simplified elasticity path diagram (van Tienderen 2000) based on our illustrative example (box 1). In this diagram, the arrows indicate how a proportional change of one variable results in a proportional change of another variable. These elasticity values can be obtained either as coefficients of the corresponding linear regressions (in data-based empirical analyses) or as derivatives (in model-based theoretical analyses). The left side of the diagram represents a classical selection analysis in the spirit of Lande and colleagues (box 3; e.g., Lande 1979, Lande and Arnold 1983), as was described above. The right side represents a demographic analysis in which elasticities characterize the effect of a proportional change in fitness components or vital rates (e.g., survival and reproduction) on a population's growth rate. The latter analyses have, for example, been found useful in conservation biology, to identify critical aspects for increasing the growth rate of endangered populations (e.g., Benton and Grant 1999, Caswell 2001). By marrying these normally disparate analyses—the former being typically more oriented toward evolution, the latter more toward ecology—mean-standardized selection gradients assume an important integrative function.

This integrative function extends to a comparative dimension. In subpopulation A of our illustrative example (box 1, figure 3), the elasticity with respect to the annual survival probability of juvenile fish (0.75) is greater than that with respect to the other fitness components (0.25), which indicates that the former is the most influential determinant of this subpopulation's growth. This is in contrast to the analogous situation in subpopulation B. Likewise, we see that the effect of the annual juvenile growth increment on newborn and juvenile survival is slightly weaker in subpopulation A (−0.71) than in subpopulation B (−0.83). Joining the two sides of the diagram, we can calculate the elasticities of the population growth rate with respect to the annual juvenile growth increment (0.01) and the date of river entry (0.23) for subpopulation A, which tells us

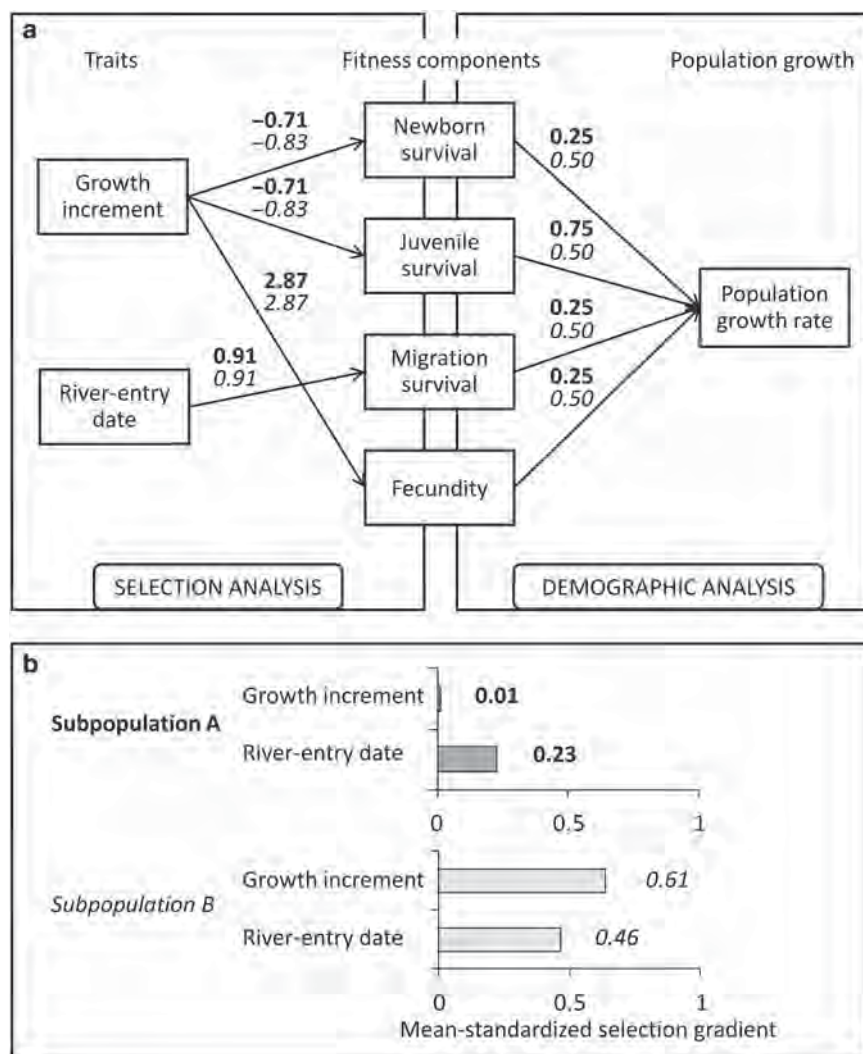


Figure 3. Elasticity path diagram and its comparative application. (a) Elasticity path diagram for the pairwise relationships between two phenotypic traits, four fitness components, and the population growth rate in subpopulations A and B of the illustrative example described in box 1. The arrows indicate how a proportional change of one variable results in a proportional change of another variable. Elasticities are shown in bold and italics for subpopulations A and B, respectively. For example, a 100% increase in the annual juvenile growth increment results in a 287% increase in fecundity (2.87). The fitness elasticity of a phenotypic trait is then calculated by following all left-to-right paths from the trait to the population growth rate, multiplying elasticities along the arrows and summing over all paths: In subpopulation A, the annual juvenile growth increment has a fitness elasticity of $(-0.71 \times 0.25) + (-0.71 \times 0.75) + (2.87 \times 0.25) = 0.01$, whereas the date of river entry has a fitness elasticity of $0.91 \times 0.25 = 0.23$. For further details on elasticity path diagrams, see van Tienderen (2000). (b) Comparison of mean-standardized selection gradients between subpopulations A and B. Parameters: $a_m = 4$ years (subpopulation A) or 2 years (B), $h_{max} = 60$ centimeters per year (A) or 55 centimeters per year (B), all other parameters as in figure 2. (See box 1 for details.)

how an increase in these phenotypic trait values results in an increase in the population growth rate. The corresponding values for subpopulation B (0.61 and 0.46, respectively)

are very different—considerably larger. Importantly, these elasticities of fitness with respect to the phenotypic traits are nothing other than mean-standardized selection gradients, because the population growth rate can be interpreted as fitness. In other words, if a small change in a trait could enhance population growth rate greatly, it also implies that selection acting on that trait is strong. Therefore, elasticities—that is, mean-standardized values—work as versatile common currencies at the interface of ecological and evolutionary dynamics, in a way that is difficult to achieve through other techniques. In fact, the elasticity path diagram has played an important role as a gateway for researchers to become familiar with the mean-standardized selection gradient (box 4). This perspective has great potential to facilitate comparisons across traits, populations, and species (box 1, figure 3) and to integrate analyses of ecological and evolutionary dynamics (Lande 1982, van Tienderen 2000) in natural populations.

Conclusions

Because the selection differential (S) and the selection gradient (β) are not dimensionless, they must be standardized before the execution of any kind of comparison of selection strengths across traits, populations, or species. We believe that biologists will benefit from routinely using the mean-standardized selection gradient in future studies. The reason for recommending this particular standardization is fundamental: It is the most direct measure of the selective implications of particular ecological settings in nature and, as such, is not confounded by a selected trait's phenotypic variability in the population under consideration.

Figure 4 summarizes the systematic standardization process that translates what can be measured in natural populations to the dimensionless measure most suitable for comparing selection strengths among traits, populations,

and species. We start our overview from what is probably the most directly observable quantity informing us about the strength of selection in natural populations: a trait's

Box 3. A roadmap to estimating mean-standardized selection gradients using phenotypic data from the wild.

In studies of selection in natural populations, the mean-standardized selection gradient will most commonly be obtained from the unstandardized selection gradient estimated from a regression-based model of relative fitness in dependence on a phenotypic trait. Here, we briefly describe the procedure for estimating the mean-standardized selection gradient from phenotypic data from natural populations. We focus on the standardization processes; the general principles of regression-based selection estimation can be found elsewhere (e.g., Brodie et al. 1995). We hope that this outline will help empirical ecologists who wish to apply the mean-standardized selection gradient to their own data.

Generally, investigators will have to account for four related steps when attempting to arrive at mean-standardized selection gradients through regression models:

Collection of empirical data. Estimation of the selection gradient from empirical observations using regression models requires individual-level data on phenotypic traits and the corresponding fitness. Although lifetime measures of fitness are ideal, in most empirical studies, only the components of an individual's fitness can be empirically assessed (e.g., in a marine fish with external broadcast reproduction, it would be impossible to estimate reproductive output by screening all potential offspring using genetic methods). Typical measures of fitness components include fertility and fecundity (e.g., the number or biomass of offspring), survival at a given stage, and mating success (Brodie et al. 1995).

Regression. The type of regression analysis depends on the structure of the chosen fitness measure. In most cases, univariate or multivariate linear regression models will be used when the fitness measure is quantitative. By contrast, logistic regression models will be suitable for binary fitness measures (such as survival; Carlson et al. 2007). Note that in the latter case, the parameter estimate for the logistic regression coefficient will need to be transformed to its linear equivalent before standardization of selection strength (Janzen and Stern 1998).

Removal of nongenetic environmental effects. If phenotypic data are available in time series, any salient environmental variables (e.g., biomasses or densities of the focal species and its potential prey, predators, or competitors or relevant abiotic variables) can be included in the regression model to control for their effects on relative fitness experienced by an individual in dependence on its phenotypic trait and varying environmental conditions.

Standardization. The unstandardized selection gradient is obtained as a coefficient of the regression of relative fitness on the phenotypic trait (or as a partial regression coefficient in the case of multivariate analyses). To obtain the mean-standardized selection gradient, the unstandardized selection gradient needs to be multiplied by the mean of the phenotypic trait in the population (equation 6). In some applications, the trait values are standardized to a mean of 0 and an SD of 1 (e.g., through z-transformation) prior to the regression analysis (e.g., Carlson et al. 2007). In this case, the regression coefficient obtained represents the SD-standardized selection gradient. Therefore, it must be multiplied by the original mean and divided by the original SD of the phenotypic trait to obtain the mean-standardized selection gradient (equations 5 and 6). Fitness measures are often generation based, in which case the selection gradient obtained from a regression analysis need not be standardized by generation time. Exceptions occur when fitness is measured in a time unit other than generation length (e.g., annual population growth rates; Arlinghaus et al. 2009), in which case multiplication with generation length is required.

evolutionary rate—the change in a phenotypic trait value divided by the time it took for the change to occur. Such evolutionary rates have been used to compare the pace of evolutionary changes in response to particular (often anthropogenic) selection pressures affecting specific populations and specific traits (e.g., Jørgensen et al. 2007, Darimont et al. 2009). Despite their accessibility to measurement, these rates, as such, however, cannot serve as suitable measures of selection strength. Although this conclusion is formally evident from the fact that such rates are not dimensionless, we can also understand it by appreciating that evolutionary rates confound the strength of selection with properties such as generation time, heritability, phenotypic variance, and phenotypic mean (figure 4), which all idiosyncratically depend on the populations and traits under consideration. It is therefore easy to see that the effects of these

population- and trait-specific confounding factors must be removed before comparisons of selection strengths among traits, populations, and species can become meaningful. As is shown in figure 4, it is the mean-standardized selection gradient (β_{μ}) that remains once the aforementioned confounding factors have been removed. In the figure, we show the steps from the evolutionary rate—the most directly observable quantity—to the dimensionless quantity of the mean-standardized selection gradient, in what to us seems the most easily understandable order. We emphasize that the order of standardization is arbitrary, which implies that quantities other than those shown in the figure can be used as stepping stones along the way. Figure 4 also reminds us of the importance of standardizing selection strengths by generation time: If a selection gradient is calculated on the basis of a population's intrinsic growth rate (Lande 1982,

Box 4. Is the mean-standardized selection gradient being increasingly used?

As was described in the main text, the mean-standardized selection gradient has some fundamental advantages as a measure of the strength of natural selection, relative to the standard deviation (SD)—standardized selection gradient. In particular, it does not confound selection strength with phenotypic variability. It is therefore interesting to evaluate the uptake and current prevalence of this measure in the relevant scientific literature. To this end, we carried out a literature survey based on the Thomson Reuters Web of Science. First, we chose five primary journal articles in which the definition of mean-standardized selection gradients and their calculation were discussed (Morgan and Schoen 1997, van Tienderen 2000, Hansen et al. 2003, Hereford et al. 2004, Stinchcombe 2005). Second, we assembled a list of secondary journal articles included in the Web of Science by the end of March 2011 in which the primary articles were cited. We identified 179 papers (excluding the 5 primary articles), which in a third step, we finally assessed for their usage of different methods for standardizing selection strength.

Surprisingly, among these 179 studies, there were only 11 in which empirical estimates of the mean-standardized selection gradient were reported (table 1). In three of these papers, the mean-standardized selection gradient was calculated in the context of elasticity path diagrams (van Tienderen 2000). This suggests that the elasticity analysis of Leslie-matrix models has played an important role as a gateway for a wider range of biologists to become familiar with the mean-standardized selection gradient. We also found that, except for Arlinghaus and colleagues (2009), in not a single model-based analysis published before the end of March 2011 for estimating selection strengths across a range of scenarios was the mean-standardized selection gradient used. Most important, the results of our survey show that the standardization of selection gradients by a population's phenotypic mean is as yet far away from becoming common practice, despite this method's superior suitability for comparing selection strengths in natural populations. However, our results also indicate that there is a small core group of authors who have increased their use of the mean-standardized selection gradient during the last few years (table 1). Moreover, some researchers (e.g., Mojica and Kelly 2010) have started to report both the SD-standardized and the mean-standardized selection gradients, as was recommended by Hereford and colleagues (2004) to enable various types of comparisons in future meta-analyses.

Why is the use of the mean-standardized selection gradient still infrequent in the contemporary literature? A reason might be gleaned from Seamons and colleagues (2007), in which the mean-standardized selection gradient was calculated but not reported, because the authors thought the values that they had obtained were unrealistic. In their own words, “[The mean-standardized selection gradients] calculated for [spawner body] length were unrealistic (much larger than unity), so the results of this analysis are not shown” (p. 414). This finding might be related to the third point mentioned by Kingsolver and Pfennig (2007)—that large mean-standardized selection gradients are observed in association with small phenotypic coefficients of variation. Although a statistical explanation has been suggested for explaining this pattern (Kingsolver and Pfennig 2007), further investigation is certainly required. It is also possible, however, that scientists are liable to misinterpret the comparison between mean-standardized selection gradients for fitness (which equals 1 by definition) and other traits, because nothing at all is a priori wrong if the latter exceed 1.

Through our literature survey, we also encountered an almost Babylonian confusion of terminology. The fact that the two common measures of standardized selection strength have been used by different authors in different contexts has increased the number of terms in circulation to a level that we consider problematic (table 2). The term *intensity of selection* (or simply *selection intensity*) has long been established in quantitative genetics for referring to the SD-standardized selection gradient—in particular, in the context of truncation selection. Since this measure was also simply called the *standardized selection differential* in a popular textbook (Falconer and Mackey 1996), several authors have used this rather unspecific term in their articles. In studies of selection on multivariate traits, following Lande and Arnold (1983), the term *standardized selection gradient* has been widely employed. The same measure, however, is sometimes also simply called the *selection gradient* (e.g., Kingsolver et al. 2001), because standardization by the phenotypic SD has been—and still is—so common. Given such defaults, the terminological situation inevitably became messy when an alternative approach to standardization—the use of units of the phenotypic mean instead of the phenotypic SD—was introduced to the field.

In the future, the two standardized measures of selection strength need to be clearly distinguished through appropriate terminology. Recently, the pairing of *variance-standardized selection gradient* and *mean-standardized selection gradient* seems to be becoming popular in review papers and meta-analyses (e.g., Hereford et al. 2004, Stinchcombe 2005, Kingsolver and Pfennig 2007). Strictly speaking, however, the selection gradient in the former case is standardized by the phenotypic SD and not by the phenotypic variance. Terms adequately reflecting this could be *selection gradient in units of the phenotypic SD* and *selection gradient in units of the phenotypic mean*. The corresponding abbreviated terms could be *SD-standardized selection gradient* and *mean-standardized selection gradient*, which is the convention we have adopted throughout the present article.

Finally, we must emphasize the importance of better distinguishing the (unstandardized) selection differential and the selection gradient. In the common regression-based approach to estimating selection from empirical data, the selection differential is the covariance between relative fitness and the trait of interest (its unit is [trait]), whereas the selection gradient is a regression coefficient of relative fitness on the trait—that is, the covariance:variance ratio of that trait (its unit is [trait⁻¹]; Lande and Arnold 1983). Yet, we noticed that the term *selection differential* was used for regression coefficients when univariate regressions of relative fitness on one trait were applied, whereas the term *selection gradient* was used in multivariate regressions jointly including several adaptive traits (e.g., Kingsolver et al. 2001, Carlson et al. 2007). This confusion might have arisen because both the selection differential and a regression coefficient in a univariate regression reflect changes resulting from a direct link between fitness and the trait (direct selection), as well as from correlations with other traits affecting fitness (indirect selection). However, a regression coefficient is not identical to the selection differential defined in traditional quantitative genetics. To avoid further confusion, we propose that the term *selection gradient* always be used when referring to coefficients of regressions of relative fitness on phenotypic traits.

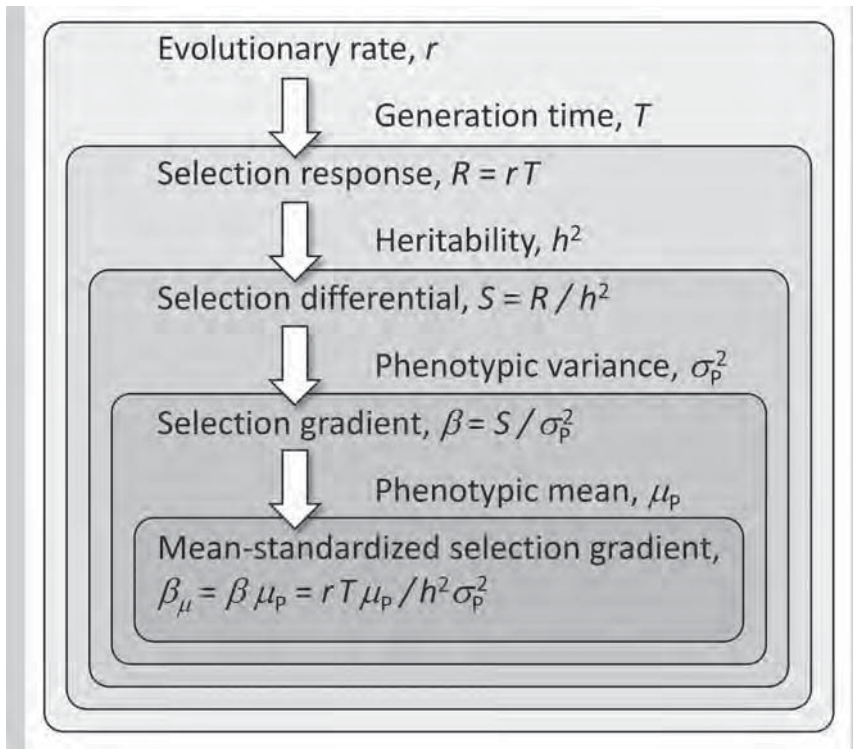


Figure 4. Successively stripping away four population- and trait-specific properties translates evolutionary rates into mean-standardized selection gradients. This standardization process can be likened to the peeling of an onion, with the directly observable quantity as the outer skin and the dimensionless quantity most useful for comparisons across traits, populations, and species at the core.

van Tienderen 2000), the result must still be standardized by the population's generation time before using it for comparing selection strengths across populations in a manner that is independent of their generation times. Box 3 outlines the most important issues to consider in regression-based analyses of empirical data for determining mean-standardized selection gradients.

Unfortunately, for historical reasons, the number of studies in which the mean-standardized selection gradient was reported is relatively small (box 4). On the basis of this observation, we would like to repeat Hereford and colleagues' (2004) appeal: Authors, reviewers, and journal editors should insist on the publication of basic yet fundamental values in studies on selection strengths, including phenotypic means and variances. If phenotypic means and variances are available, any measure of selection strength, reported as a selection differential (S) or as a selection gradient (β), can easily be converted into the mean-standardized selection gradient (figure 4, equation 6). It is only under this condition that reported selection strengths become comparable across traits, populations, and species and therefore become amenable to future meta-analyses.

To conclude, we agree with Hereford and colleagues' (2004) caveat that no standardized measure of selection strength

will be satisfactory for all research purposes. The reason for this is conceptually profound and is intimately related to the notion of a trait's natural scale and measurement theory (box 2; Houle et al. 2011). At any rate, jointly reporting unstandardized, SD-standardized, and mean-standardized selection gradients in all salient scientific publications will allow readers to conduct their own analyses and to draw their own conclusions (Stinchcombe 2005) and is therefore the recommended way forward.

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